


Matrigel Invasion Assay Protocol

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Derivation of trophoblast stem cells from naïve human pluripotent stem cells

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Detailed protocol

Materials and Reagents

1. EVTs or hTSCs
2. EVT basal medium [DMEM/F12 (Gibco, 11320) supplemented with 0.1 mM β -mercaptoethanol (Millipore Sigma, 8.05740), 0.5% penicillin-streptomycin (Gibco, 15140), 0.3% BSA (Gibco, 15260), 1% ITS-X (Gibco, 51500), 7.5 mM A83-01 (BioVision, 1725), 2.5 mM Y27632 (Stemgent, 040012)]
3. TSC medium [DMEM/F12 supplemented with 0.1 mM β -mercaptoethanol, 0.2% FBS, 0.5% Penicillin-Streptomycin, 0.3% BSA, 1% ITS-X, 1.5 mg/ml L-ascorbic acid (Wako, 013-12061), 50 ng/ml EGF (Rockland, 009-001 C26), 2 mM CHIR99021 (Stemgent, 04-0004), 0.5 mM A83-01, 1 mM SB431542 (BioVision, 1674), 0.8 mM VPA (Tocris, 2815), and 5 mM Y-27632]
4. Fetal bovine serum (FBS) (Millipore Sigma, ES-009-B)
5. TrypLE Express (Gibco, 12604)
6. Matrigel-coated transwell inserts with 8.0 μ m pores (Corning, 354480)
7. 4% PFA
8. Cotton tipped swab
9. PBS

Equipment

1. Centrifuges
2. Humidified tissue culture incubator, 37°C, 5% CO₂
3. Microscope

Procedure

1. Preparation of trans-well inserts

- a. Place desired number of matrigel-coated inserts into a 24-well companion plate and allow them to come to room temperature.
- b. Add warm (37°C) bicarbonate based culture medium to the interior of the inserts and bottom of wells. Allow the inserts to rehydrate for 2 hours in a humidified tissue culture incubator.
- c. After rehydration, carefully remove the medium without disturbing the layer of Corning® Matrigel® Matrix on the membrane. Don't let the Matrigel dry, so make sure the cells are ready when the Matrigel is rehydrating.

2. Prepare cells (Upper Chamber)

- a. Rinse EVTs or hTSCs once with 10 mL PBS; add 3 mL of TrypLE Express and incubate at 37°C for 5 min; neutralize the trypsin with FBS containing EVT/TSC medium.
- b. In a 50 mL conical tube, centrifuge cells at 300 \times g for 10 min.
- c. Count cells using a hemocytometer.
- d. Prepare 2.0×10^5 cells in 200 μ L of EVT basal medium or TSC medium.
- e. Cap tube and store at room temperature until they are ready to be loaded in the upper chamber of transwell inserts.

3. Prepare the chemoattractant (For Bottom Chamber)

- a. For the bottom chamber, prepare 800 μ L of EVT basal medium or TSC medium supplemented with 20% FBS for each well.
- b. Add 800 μ L of chemoattractant carefully to each well. Avoid creating bubbles.

4. Assemble the Invasion Chamber

- a. Using sterile forceps, carefully transfer the Matrigel-coated inserts to the wells containing chemoattractant. Make sure there are no air bubbles trapped beneath the insert. If there are bubbles, remove insert and try again.
- b. Add 200 μ L of cell suspension (preparation as described above) to the upper chamber of Matrigel-coated inserts.
- c. Incubate the inserts at 37°C, 5% CO₂ for 36 h for Cell Invasion Assay.

5. Measurement of cell invasion

- a. After invasion period, carefully aspirate the media from the insert without disturbing the Matrigel layer and wash twice with PBS.
- b. Fix inserts with 500 μ L of 4% paraformaldehyde for 15 min in an empty well.
- c. Remove fixing solution and wash twice with PBS.
- d. Add 500 μ L of crystal-violet solution to the empty wells and place the inserts over it for 15 min to stain the cells.
- e. Remove inserts from the crystal-violet solution and rinse inserts in three small beakers containing PBS to remove the extra stain.
- f. Remove the non-invaded cells on the upper chamber carefully by scrubbing with a cotton tipped swab. Repeat the scrubbing with a second swab moistened with medium.
- g. Place the insert on a slide and count the invasive cells under a microscope and capture the images.

References

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